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18 DEC 2000 CT

09/720066

Certificate of Mailing

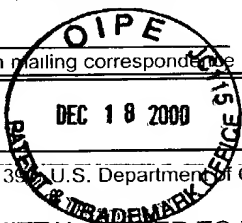
Date of Deposit December 18, 2000Label Number: EL509218366US

I hereby certify under 37 CFR 1.10 that this correspondence is being deposited with the United States Postal Service as "Express Mail Post Office to Addressee" with sufficient postage on the date indicated above and is addressed to BOX PATENT APPLICATION, Assistant Commissioner of Patents, Washington, D.C. 20231

Guy Beardsley

Printed name of person mailing correspondence

Signature of person mailing correspondence



Substitute Form PTO 1350 U.S. Department of Commerce Patent and Trademark Office

Attorney's Docket Number:
50125/019001

TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371

U.S. Application Number

09/720066

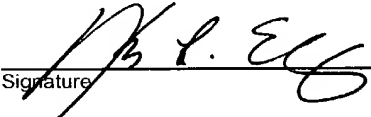
INTERNATIONAL APPLICATION NUMBER		INTERNATIONAL FILING DATE	PRIORITY DATE CLAIMED
PCT/EP99/04288		June 21, 1999 (21.06.98)	June 19, 1998 (19.06.98)
TITLE OF INVENTION:		AAV STRUCTURAL PROTEIN, ITS PREPARATION AND USE	
APPLICANTS FOR DO/EO/US:		Michael Hallek et al.	
Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information:			
1.	X	This is a FIRST submission of items concerning a filing under 35 U.S.C. 371.	
2.		This is a SECOND or SUBSEQUENT submission of items concerning a filing under 35 U.S.C. 371.	
3.	X	This is an express request to begin national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b) and PCT Articles 22 and 39(1).	
4.	X	A proper Demand for International Preliminary Examination was made by the 19 th month from the earliest claimed priority date.	
5.		A copy of the International Application as filed (35 U.S.C. 371(c)(2)).	
a.		is transmitted herewith (required only if not transmitted by the International Bureau).	
b.		has been transmitted by the International Bureau.	
c.		Is not required, as the application was filed with the United States Receiving Office (RO/US).	
6.	X	A translation of the International Application into English (35 U.S.C. 371(c)(2)).	
7.		Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371(c)(3)).	
a.		are transmitted herewith (required only if not transmitted by the International Bureau).	
b.		have been transmitted by the International Bureau.	
c.		have not been made; however, the time limit for making such amendments has NOT expired.	
d.		have not been made and will not be made.	
8.	X	A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371(c)(3)).	
9.	X	An oath or declaration of the inventors (35 U.S.C. 371(c)(4)).	
10.		A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371(c)(5)).	

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11.		An Information Disclosure Statement under 37 CFR 1.97 and 1.98.		
12.		An assignment for recording. A separate cover sheet in compliance with 37 3.28 and 3.31 is included.		
13.	X	A FIRST preliminary amendment.		
		A SECOND or SUBSEQUENT preliminary amendment.		
14.		A substitute specification.		
15.		A change of power of attorney and/or address letter.		
16.		Other items or information:		
17.	X	The following fees are submitted: BASIC NATIONAL FEE (37 CFR 1.492(A)(1)-(5)): Search Report has been prepared by the EPO or JPO \$ 860.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) \$ 690.00 No international preliminary examination fee paid to USPTO (37 CFR 1.482) but international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$ 710.00 Neither international preliminary examination fee (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$ 1000.00 International preliminary examination fee paid to USPTO (37 CFR 1.482) and all claims satisfied provisions of PCT Article 33(2)-(4) \$ 100.00		
		ENTER APPROPRIATE BASIC FEE AMOUNT =		\$ 1000.00
		Surcharge of \$130 for furnishing the oath or declaration later than <input type="checkbox"/> 20 OR <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(e)).		\$
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE	
Total claims	27 - 20 =	7	x \$18.00	\$ 126.00
Independent claims	1 - 3 =		x \$80.00	\$
Multiple dependent claims (if applicable)			+ \$270.00	\$ 270.00
		TOTAL OF ABOVE CALCULATIONS =		\$1396.00
SMALL ENTITY STATUS:				
Applicant claims small entity status under 37 CFR 1.27.				
Reduction of 1/2 for filing by small entity, if applicable. Verified Small Entity Statement must also be filed with this request (Note 37 CFR 1.9, 1.27, 1.28).			\$ 698.00	
SUBTOTAL =			\$ 698.00	
Processing fee of \$130.00 for furnishing the English translation later than <input type="checkbox"/> 20 OR <input type="checkbox"/> 30 months from the earliest claimed priority date (37 CFR 1.492(f)).			+	\$
TOTAL NATIONAL FEE =			\$	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31). \$40.00 per property.			+	\$
TOTAL FEES ENCLOSED =			\$ 698.00	

09/17/2000 66'
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		Amount to be refunded	\$
		charged	\$
a.	X	A check in the amount of \$698.00 to cover the above fees is enclosed.	
b.		Please charge my Deposit Account No. 03-2095 in the amount of \$ [**. **] to cover the above fees.	
c.	X	The Commissioner is hereby authorized to charge any additional fees which may be required, or credit any overpayment, to Deposit Account No. 03-2095. A duplicate copy of this sheet is enclosed.	
NOTE: Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (37 CFR 1.137(a) or (b)) must be filed and granted to restore the application to pending status.			
SEND ALL CORRESPONDENCE TO:			
Karen L. Elbing, Ph.D. Clark & Elbing LLP 176 Federal Street Boston, MA 02110-2214 Telephone: 617-428-0200 Facsimile: 617-428-7045		 Signature Karen L. Elbing, Ph.D. Reg No. 35,238	

\\ntserver\documents\50125\50125.019001 U.S. Natnl. Phase Application transmittal & formal papers.wpd

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534 Rec'd PCT/PTO 18 DEC 2000

PATENT
ATTORNEY DOCKET NUMBER: 50125/019001

Certificate of Mailing	
Date of Deposit <u>December 18, 2000</u>	Label Number: <u>EL509218366US</u>
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<u>Guy Beardsley</u> Printed name of person mailing correspondence	<u>Guy Beardsley</u> Signature of person mailing correspondence

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant: Michael Hallek et al.

Art Unit:

Serial No.: Not yet assigned

Examiner:

Filed: December 18, 2000

Title: AAV STRUCTURAL PROTEIN, ITS PREPARATION AND USE

Assistant Commissioner of Patents
Washington, D.C. 20231

PRELIMINARY AMENDMENT

Prior to examination on the merits, kindly amend the above-referenced application as follows.

In the claims:

Amend claims 3, 4, 6-8, 11-14, 17, 18, and 20-23 as follows.

3. (Amended) Structural protein according to [either of Claims 1 or 2] Claim 1, characterized in that the mutation(s) is/are located at the N terminus of the structural protein.

4. (Amended) Structural protein according to [any of Claims 1 to 3] Claim 1, characterized in that the mutated structural protein brings about a change in the protein-cell membrane receptor interaction.

6. (Amended) Structural protein according to [any of Claims 1 to 5] Claim 1, characterized in that it is selected from mutated VP1, mutated VP2 and/or mutated VP3.

7. (Amended) Structural protein according to [any of Claims 1 to 6] Claim 1, characterized in that it is derived from AAV2, AAV3, AAV4, AAV5 and/or AAV6.

8. (Amended) Structural protein according to [any of Claims 1 to 7] Claim 1, characterized in that the mutation(s) is/are point mutation(s), mutation(s) of several amino acids, one or more deletions and/or one or more insertions, or a combination of this mutation.

11. (Amended) Structural protein according to [any of Claims 8 to 10] Claim 8, characterized in that the mutation(s) is/are brought about by one or more insertions at the XhoI cleavage site of the VP1-encoding nucleic acid.

12. (Amended) Structural protein according to [any of Claims 8 to 10] Claim 8, characterized in that the mutation(s) is/are brought about by one or more insertions at the BsrBI cleavage site of the VP1-encoding nucleic acid.

13. (Amended) Structural protein according to [any of Claims 8 to 10] Claim 8, characterized in that the mutation(s) is/are brought about by one or more deletions between the BsrBI/HindII cleavage sites of the VP1-encoding nucleic acid and one or more insertions.

14. (Amended) Structural protein according to [any of Claims 8 to 10] Claim 8, characterized in that one or more insertions in VP3 is/are located before and/or after at least one amino acid in the sequence selected from YKQIS SQSGA, YLTLN NGSQA, YYLSR TNTPS, EEKFF PQSGV, NPVAT, EQYGS, LQRGN RQAAT, NVDFV VDTNG.

17. (Amended) Structural protein according to [any of Claims 1 to 16] Claim 1 in the form of an AAV particle, in particular in the form of an AAV capsid.

18. (Amended) Nucleic acid coding for a structural protein according to [any Claims 1 to 16] Claim 1.

20. (Amended) Process for the preparation of a structural protein according to [any of Claims 1 to 16] Claim 1, characterized in that a cell according to Claim 19 is cultivated and, where appropriate, the expressed structural protein is isolated.

21. (Amended) Medicinal product comprising a structural protein according to [any of Claims 1 to 17] Claim 1, a nucleic acid according to Claim 18 and/or a cell according to Claim 19.

22. (Amended) Diagnostic aid comprising a structural protein according to [any of Claims 1 to 17] Claim 1, a nucleic acid according to Claim 18 and/or a cell according to Claim 19.

$\frac{1}{2} \begin{pmatrix} 1 & 0 \\ 0 & 1 \end{pmatrix}, \frac{1}{2} \begin{pmatrix} 1 & i \\ 0 & -1 \end{pmatrix}, \frac{1}{2} \begin{pmatrix} 1 & -i \\ 0 & -1 \end{pmatrix}, \frac{1}{2} \begin{pmatrix} 1 & 0 \\ 0 & -1 \end{pmatrix}, \frac{1}{2} \begin{pmatrix} 1 & i \\ 0 & 1 \end{pmatrix}, \frac{1}{2} \begin{pmatrix} 1 & -i \\ 0 & 1 \end{pmatrix}, \frac{1}{2} \begin{pmatrix} 1 & 0 \\ 0 & 1 \end{pmatrix}, \frac{1}{2} \begin{pmatrix} 1 & i \\ 0 & -1 \end{pmatrix}, \frac{1}{2} \begin{pmatrix} 1 & -i \\ 0 & -1 \end{pmatrix}, \frac{1}{2} \begin{pmatrix} 1 & 0 \\ 0 & -1 \end{pmatrix}$

REMARKS

2095.

Respectfully submitted,

Date: 18 December 2000

~~Karen L. Elbing, Ph.D.~~
~~Reg. No. 35,238~~

Clark & Elbing LLP
176 Federal Street
Boston, MA 02110
Telephone: 617-428-0200
Facsimile: 617-428-7045

\\Ntserver\documents\50125\50125 19001 Preliminary Amendment for claims in application.wpd

PTO/PCT Rec'd 19 OCT 2001 15 30 19 01

PATENT
ATTORNEY DOCKET NO. 50125/019001

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I hereby certify under 37 C.F.R. § 1.8(a) that this correspondence is being deposited with the United States Postal Service as **first class mail** with sufficient postage on the date indicated above and is addressed to the Assistant Commissioner for Patents, Washington, D.C. 20231.

Colleen Coyne

Printed name of person mailing correspondence

Colleen Coyne

Signature of person mailing correspondence

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant:	Michael Hallek et. al.	Art Unit:	Not Yet Assigned
Serial No.:	09/720,066	Examiner:	Not Yet Assigned
Filed:	December 18, 2000	Customer No.:	21559
Title:	AAV STRUCTURAL PROTEIN, ITS PREPARATION AND USE		

Assistant Commissioner For Patents
Washington, DC 20231

PRELIMINARY AMENDMENT

Prior to examination, kindly amend the above-referenced application as follows.

In the Specification:

Insert the attached sequence listing at the end of the application.

Please replace the last paragraph on page 9 with the following paragraph re-written in clean form:

An example of a particularly preferred ligand is the P1 peptide
(QAGTFALRGDNPQG) (SEQ ID NO: 1) which is a peptide 14 amino acids long from

the core sequence of an alpha chain of the laminin family. This sequence is sufficient, for example, to recognize an integrin receptor which mediates, inter alia, the endocytosis of viral particles, for example of adenovirus. The P1 peptide binds irrespective of its conformation (linear or circular) to the integrin receptor. According to the present invention, the coding DNA sequence of the P1 peptide is inserted into the gene coding for an AAV

Please replace the first full paragraph on page 11 with the following paragraph re-written in clean form:

In a further preferred embodiment, one or more insertions are present in the VP3 structural protein (Rutledge, E.A. et al. (1998) *supra*) before and/or after at least one amino acid in the sequence selected from, YKQIS SQSGA (SEQ ID NO: 2), YLTLN NGSQA (SEQ ID NO: 3), YYLSR TNTPS (SEQ ID NO: 4), EEKFF PQSGV (SEQ ID NO: 5), NPVAT EQYGS (SEQ ID NO: 6), LQRGB RQAAT (SEQ ID NO: 7), NVDFT VDTNG (SEQ ID NO: 8), because these sites are located on the exposed sites of a loop, in which case the risk of changing the VP3 structure is low.

Please replace the first full paragraph on page 17 with the following paragraph re-written in clean form:

2. Mutations in VP3

a) ins447; YYLSR TNTPS (CPV: 300) (SEQ ID NO: 4)

b) **INS534**; **EEKFF PQSGV** (CPV: 390) (SEQ ID NO: 5)

c) **ins573**; **NPVAT EQYGS** (CPV: 426) (SEQ ID NO: 6)

d) **ins587**; **LQRGN RQAAT** (CPV: 440) (SEQ ID NO: 7)

e) **ins713**; **NVDFT VDTNG** (CPV: 565) (SEQ ID NO: 8)

CPV means here the location in the equivalent CPV capsid

(Named according to the number of amino acids (AAs) counted after the AA at the start of the N terminus in the VP-1 of AAV2, flanked by in each case 5 amino acids located N-terminally thereof and 5 amino acids located C-terminally thereof; the AA after which the insertion was introduced is shown bold).

Please replace the last paragraph on page 19 with the following paragraph re-written in clean form:

The initial starting point was a plasmid pUC-AV2 which was prepared by subcloning of the 4.8 kb BglII fragment of pAV2 (ATCC 37261, ref. 53) into the BamHI cleavage site of pUC19 (New England BioLabs Inc.). Mutations were carried out at defined sites in the plasmid by means of PCR-assisted mutagenesis known to the skilled person. This entailed a sequence coding for P1, a 14 AA peptide with the AA sequence, QAGTFALRGDNPQG (SEQ ID NO: 1), which contains the RGD binding motif of a laminin fragment (Aumailly et al. (1990) FEBS Lett. 262, 82-86), being inserted after nucleotides 3543, 3804, 3921, and 3963. This corresponds to an insertion after amino acids 447, 534, 573, and 587 of the AAV2 capsid protein (named according

to the number of amino acids (AA) counted after the AA at the start of the N terminus in VP-1 of AAV2). In the subsequent PCR there is use of in each case 2 mutation-specific primers and, as template, a plasmid, pCap, which contains only the cap gene and is formed by cutting out the 2.2 kb

In the Claims:

Amend claim 14 as follows.

14. (Twice Amended) Structural protein according to Claim 8, characterized in that one or more insertions in VP3 is/are located before and/or after at least one amino acid in the sequence selected from YKQIS SQSGA (SEQ ID NO: 2), YLTLN NGSQA (SEQ ID NO: 3), YYLSR TNTPS (SEQ ID NO: 4), EEKFF PQSGV (SEQ ID NO: 5), NPVAT, EQYGS (SEQ ID NO: 6), LQRGN RQAAT (SEQ ID NO: 7), and NVDFT VDTNG (SEQ ID NO: 8).


REMARKS

The specification has been amended to provide a unique sequence identification number for each nucleotide sequence. The attached sequence listing has also been inserted into the application. No new matter is introduced by any of these amendments.

Please apply any charges or credits to Deposit Account 03-2095.

Respectfully submitted,

Date: 15 October 2001


 Karen L. Elbing, Ph.D.
 Reg. No. 35,238

Clark & Elbing LLP
 176 Federal Street
 Boston, MA 02110
 Telephone: 617-428-0200
 Facsimile: 617-428-7045

50125.019001 Preliminary Amendment.wpd



Marked Up Version of Amended Claim

14. (Twice Amended) Structural protein according to Claim 8, characterized in that one or more insertions in VP3 is/are located before and/or after at least one amino acid in the sequence selected from YKQIS SQSGA (SEQ ID NO: 2), YLTLN NGSQA (SEQ ID NO: 3), YYLSR TNTPS (SEQ ID NO: 4), EEKFF PQSGV (SEQ ID NO: 5), NPVAT, EQYGS (SEQ ID NO: 6), LQRGN RQAAT (SEQ ID NO: 7), and NVDFTVDTNG (SEQ ID NO: 8).

09720066, 1001901

PATENT Rec'd 19 OCT 2001

PATENT
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Colleen Coyne
Printed name of person mailing correspondence

Colleen Coyne
Signature of person mailing correspondence

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

Applicant:	Michael Hallek et. al.	Art Unit:
Serial No.:	09/720,066	Examiner:
Filed:	December 18, 2000	Customer No.: 21559
Title:	AAV STRUCTURAL PROTEIN, ITS PREPARATION AND USE	

Assistant Commissioner For Patents
Washington, DC 20231

STATEMENT UNDER 37 C.F.R. § 1.825

In reply to the Notice to Comply with Sequence Requirements mailed July 13, 2001 and as required by 37 C.F.R. § 1.825(a), enclosed is an amended sequence listing consisting of 3 sheets to be inserted at the end of the application.

The amendments correct typographical errors. I hereby submit that the substitute sheets contain no new matter.

As required by 37 C.F.R. § 1.825(b), enclosed is a diskette containing a copy of the sequence listing in computer readable form including all previously submitted data with the amendments incorporated therein. The contents of the computer readable form are the same as the contents of the paper sheets.

[illegible]

Date: 15 October 2001

~~Karen L. Elbing, Ph.D.~~

Reg. No. 35,238

Clark & Elbing LLP
176 Federal Street
Boston, MA 02110
Telephone: 617-428-0200
Facsimile: 617-428-7045



\\Clark-w2k1\documents\50125\50125.019001 Sequence Statement.wpd

SEQUENCE LISTING

<110> Hallek, Michael
 Ried, Martin
 Deleage, Gilbert
 Girod, Anne

<120> AAV Structural Protein, Its Preparation
 and Use

<130> 50125/019001

<140> US 09/720,066
 <141> 2000-12-18

<150> PCT/EP99/04288
 <151> 1999-06-21

<150> DE 19827457.2
 <151> 1998-06-19

<160> 8

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<210> 3
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 <223> Mutated adeno-associated virus VP3 protein



PCT WELTORGANISATION FÜR GEISTIGES EIGENTUM
Internationales Büro
INTERNATIONALE ANMELDUNG VERÖFFENTLICHT NACH DEM VERTRAG ÜBER DIE
INTERNATIONALE ZUSAMMENARBEIT AUF DEM GEBIET DES PATENTWESENS (PCT)

(51) Internationale Patentklassifikation ⁶ : C12N 15/35, 15/62, C07K 14/015, C12N 5/10, A61K 39/23, 48/00, G01N 33/68, C12Q 1/68	A2	(11) Internationale Veröffentlichungsnummer: WO 99/67393 (43) Internationales Veröffentlichungsdatum: 29. Dezember 1999 (29.12.99)
(21) Internationales Aktenzeichen: PCT/EP99/04288 (22) Internationales Anmeldedatum: 21. Juni 1999 (21.06.99) (30) Prioritätsdaten: 198 27 457.2 19. Juni 1998 (19.06.98) DE (71) Anmelder (für alle Bestimmungsstaaten ausser US): MEDIGENE AKTIENGESELLSCHAFT [DE/DE]; Lochhamer Strasse 11, D-82152 Martinsried (DE). (72) Erfinder; und (75) Erfinder/Anmelder (nur für US): HALLEK, Michael [DE/DE]; Brunnenstrasse 40, D-89638 Schondorf (DE). RIED, Martin [DE/DE]; Am Lohwald 36, D-86697 Sinning (DE). DELEAGE, Gilbert [FR/FR]; 43, rue Saint Romain, F-69008 Lyon (FR). GIROD, Anne [FR/DE]; Appenzeller Strasse 123, D-81475 München (DE). (74) Anwalt: BÖSL, Raphael; Bardehle, Pagenberg, Dost, Altenburg, Geissler, Isenbruck, Galileiplatz 1, D-81679 München (DE).		(81) Bestimmungsstaaten: AU, CA, JP, US, europäisches Patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). Veröffentlicht <i>Ohne internationalen Recherchenbericht und erneut zu veröffentlichen nach Erhalt des Berichts.</i>
(54) Title: AAV SCLEROPROTEIN, PRODUCTION AND USE THEREOF (54) Bezeichnung: STRUKTURPROTEIN VON AAV, SEINE HERSTELLUNG UND VERWENDUNG (57) Abstract The invention relates to an AAV (Adeno-associated virus) scleroprotein, containing at least one mutation that increases the infectiousness of the virus. (57) Zusammenfassung Die Erfindung betrifft ein Strukturprotein von Adeno-assoziiertem Virus (AAV), das mindestens eine Mutation enthält, die eine Erhöhung der Infektiosität des Virus bewirkt.		

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December 11, 2000

5 **AAV structural protein, its preparation and use**

10 The present invention relates to a structural protein of adeno-associated virus (AAV) which comprises at least one mutation which brings about an increase in the infectivity.

15 The AAV virus belongs to the family of parvoviruses. These are distinguished by an icosahedral, non-enveloped capsid which has a diameter of 18 to 30 nm and which contains a linear, single-stranded DNA of about 5 kb. Efficient replication of AAV requires coinfection of the host cell with helper viruses, for example with adenoviruses, herpesviruses or vaccinia viruses. In the absence of a helper virus, AAV enters a latent state, the viral genome being capable of stable integration into the host cell genome. The property of AAV integrating into the host genome makes it particularly interesting as a transduction vector for mammalian cells. In general, the two inverted terminal repeats (ITR) which are about 145 bp long are sufficient for the vector functions. They carry the "cis" signals necessary for replication, packaging and integration into the host cell genome. For packaging in recombinant vector particles, a vector plasmid which carries the genes for nonstructural proteins (Rep proteins) and for structural proteins (Cap proteins) is transfected into cells suitable for packaging, for example HeLa or 293 cells, which are then infected, for example, with adenovirus. A lysate containing recombinant AAV particles is obtained after some days.

20 The AAV capsid consists of three different proteins: VP1, VP2 and VP3, whose relative proportions are 5% VP1, 5% VP2 and 90% VP3. The AAV capsid genes are

- 2 -

located at the right-hand end of the AAV genome and are encoded by overlapping sequences of the same open reading frame (ORF) using different start codons. The VP1 gene contains the whole VP2 gene sequence, which in turn contains the whole VP3 gene sequence with a specific N-terminal region. The fact that the overlapping reading frames code for all three AAV capsid proteins is responsible for the obligatory expression of all capsid proteins, although to different extents.

The molecular masses of the capsid proteins are 87 kD for VP1, 73 kD for VP2 and 62 kD for VP3. The sequences of the capsid genes are described, for example, in Srivastava, A. et al. (1983), J. Virol., 45, 555-564; Muzyczka, N. (1992), Curr. Top. Micro. Immunol., 158, 97-129, Ruffing, N. et al. (1992), J. Virol., 66, 6922-6930 or Rutledge, E. A. et al. (1998) J. Virol. 72, 309-319. The physical and genetic map of the AAV genome is described, for example, in Kotin, R.M. (1994), Human Gene Therapy, 5, 793-801.

Also known are various AAV serotypes, of which the human AAV serotype 2 (AAV2) represents a virus vector with advantageous properties for somatic gene therapy. The essential advantages are the lack of pathogenicity for humans, the stable integration of viral DNA into the cellular genome, the ability to infect non-dividing cells, the stability of the virion, which makes purification to high titres (10^{11} particles per ml) possible, the low immunogenicity, and the substantial absence of viral genes and gene products in the recombinant AAV vector, which is advantageous from the viewpoint of safety for use in gene therapy. The cloning of genes into the AAV vector now takes place by methods generally known to the skilled person, as described, for example, in WO 95/23 867, in Chiorini J.A. et al. (1995), Human Gene Therapy, 6, 1531-1541 or in Kotin, R.M. (1994), supra.

AAV2 for example has in general a broad active spectrum. Epithelial tissues, such as human epithelial tumour cell lines, but also primary tumour material
 5 such as cervical or ovarian carcinoma or melanoma, and human keratinocytes are infected very efficiently (70-80%), whereas haematopoietic cells such as lymphohaemopoietic cells are infected with 10- to 100-fold lower efficiency (0.5-5%) (Mass et al. (1998)
 10 Human Gene Therapy, 9, 1049-1059). One reason for this might be that an interaction between AAV and an AAV receptor on the surface of the cell is necessary for uptake of AAV into the cell. Thus, for example, the putative primary AAV2 receptor is a cell membrane
 15 glycoprotein of 150 kD (Mizukami, H. et al. (1996), Virology, 217, 124-130) or heparan sulphate proteoglycan (Summerford, C. & Samulski, R.J. (1998), J. Virol., 72, 1438-1445). Possible secondary receptors which have been determined are: $\alpha V\beta 5$ integrin
 20 (Summerford et al., (1999) Nature Medicine 5, 78-82) and human fibroblast growth factor receptor 1 (Qing et al., (1999) Nature Medicine 5, 71-77). Binding studies have now shown that the surface density of this receptor is reduced on cells which are inefficiently
 25 infected by AAV2.

It is now known that it is possible to by genetic modification of capsid proteins of retroviruses and adenoviruses to introduce binding sites for receptors
 30 which are expressed only on particular cells into a capsid, and thus a receptor-mediated targeting of vectors has been made possible (see, for example, Cosset, F.L. & Russell, S.J. (1996), Gene Ther., 3, 946-956, Douglas, J.T. et al. (1996), Nat. Biotechnol.,
 35 14, 1574-1578, Krasnykh, V.N. et al. (1996), J. Virol., 70, 6839-6846, Stevenson, S.C. et al. (1997), J. Virol., 71, 4782-4790 or Wickman, T. J. et al. (1996), Nat. Biotechnol., 14, 1570-1573). WO 96/00587 also refers to AAV capsid fusion proteins which are said to

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contain heterologous epitopes of clinically relevant antigens, which is said to induce an immune response, and which are said not to interfere with capsid formation. However, the publication contains only a general reference without detailed information on the implementability, in particular on suitable insertion sites. Steinbach et al. (1997) (Biol. Abstr. 104, Ref. 46570) were concerned with the in vitro assembly of AAV particles which had previously been expressed in the baculo system. Mutations are also made on the cap gene, but these are intended not to lead to a change in the tropism but to a plasmid construct in which only one VP protein is expressed in each case. There is no mention of a change in the infectivity. Ruffing et al. (1994) (J. Gen. Virol. 75, 3385-3392) intended to investigate the natural tropism of AAV2. For this purpose, mutations were introduced at the C terminus of the AAV2 VP protein, the basic assumption (erroneous due to incorrect initial data) being to change an RGD motif in this way. The mutation merely brought about reduced infectivity.

Indirect targeting is disclosed in Bartlett et al. (1999; Nat. Biotechnol. 17, 181-186). In this case, there was use of a bispecific antibody which was directed both against the AAV2 capsid and against a target cell. The viral capsid was, however, neither covalently linked nor modified or a capsid protein mutated. The only attempt to date at direct targeting in the case of AAV2 was undertaken by Yang et al. (1998; Hum. Gene Ther. 1, 1929-1937). In this case, single-chain antibody fragments against the CD34 molecule was fused to the N terminus of VP2, inserted directly at the N terminus of VP1. This method has, however, 2 distinct disadvantages. On the one hand, the infection titre was very low and, on the other hand, for successful packaging it was necessary to coexpress the fusion protein with unmutated capsid proteins VP1, VP2 and VP3. However, this resulted in a mixture of

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chimeric and wild-type capsid proteins, whose composition and thus activity was unpredictable. Moreover, the packaging efficiency and the infectivity via the wild-type receptor of HeLa cells was also
5 considerably reduced compared with the wild type.

One object of the present invention was therefore to modify AAV in such a way that a more specific and more efficient gene transfer is possible than with known AAV
10 vectors.

It has now been found, surprisingly, that structural or capsid proteins of AAV can be modified so that this brings about an increase in infectivity.
15

One aspect of the present invention is therefore an AAV structural protein which comprises at least one mutation which brings about an increase in the infectivity. It is possible through the increase in
20 infectivity for example to achieve a specific and efficient gene transfer of slightly infected tissue such as, for example, haematopoietic tissue. Changing and, in particular, increasing mean for the purpose of this invention not a general but a cell-specific change
25 or increase, that is to say in relation to a particular cell type. Hence, cases in which the infectivity is reduced for particular cells and is increased only for another cell type or several other cell types are also included under an increase in the infectivity.

30 The mutation(s) is/are preferably located on the virus surface. For determining the surface-located regions of the structural proteins, it was surprisingly found according to the present invention that CPV and AAV2
35 sequences and structures are comparable. It is therefore possible to have recourse preferably to known crystal structures of parvoviruses such as of parvovirus B19 or of CPV (canine parvovirus) and to identify, with the aid of homology comparisons, protein

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domains which are important for the AAV/AAV receptor interaction and which can be modified. According to the present invention, therefore, for example a computer-assisted comparison between CPV and AAV2, and parvovirus B19 and AAV2, have surprisingly led reproducibly to the identification of loops in VP3, whose sequence varies, i.e. which have a low homology and which may be responsible for the tropism and the differences in infectivity of the virus. Thus, the known crystal structure of the CPV VP2 capsid protein (for example Luo M. (1988), J. Mol. Biol., 200, 209-211; Wu and Rossmann (1993), J. Mol. Biol., 233, 231-244) was taken as pattern, because of the great similarity to AAV2 VP3 in the secondary structure of the protein, in order to find the regions which are exposed on the viral capsid surface and, because of the local amino acid sequence, are sufficiently flexible to withstand insertion of a peptide sequence. In this case, care was taken that no secondary structural elements of the AAV2 capsid protein which would destabilize the capsid were selected.

Another possibility for determining the surface-located regions of the structural proteins is to compare the
25 nucleic acid sequences coding for the capsids from different AAV serotypes. It is possible to use for this purpose, for example, known DNA sequences from different AAV serotypes, such as AAV2, AAV3, AAV4 or AAV6, for structural analyses of possible capsid
30 morphologies of, for example, AAV2, it being possible ab initio to calculate possible tertiary structures and assign sequence regions on the basis of generally known amino acid properties to the inner or outer capsid regions. It was thus possible, for example, according
35 to the present invention to establish seven possible insertion sites in the VP3 region of the AAV2 capsid, and these made it possible to insert, for example, a ligand and express it on the viral surface (see below).

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In another preferred embodiment, the mutation(s) are located at the N terminus of the structural protein, because it has been found that, for example, in the case of the parvoviruses CPV and B19 the N terminus is located on the cell surface. In this case, the mutation is preferably not carried out directly at the N terminus of VP1 but is carried out a few amino acids downstream from the N terminus.

10 In another preferred embodiment, the mutation causes a
change in the protein-cell membrane receptor
interaction, the cell membrane receptor preferably
being a glycoprotein of about 150 kD and/or a heparan
sulphate proteoglycan, as described above in detail.
15 These two receptors are presumably primary receptors
which are supplemented by at least one secondary
receptor (see above).

In general, the mutation may be present in the VP1, VP2 and/or VP3 structural protein, with the VP1 structural protein and/or the VP3 structural protein being preferred. The mutated structural protein is furthermore preferably still capable of particle formation, i.e. formation of an icosahedral capsid. The structural protein may furthermore be derived from all AAV serotypes, in particular from human serotypes, preferably from AAV1, AAV2, AAV3, AAV4, AAV5 and/or AAV6, especially from AAV2, AAV3 and/or AAV6. These also include serotypes derived from said serotypes, in particular AAV2.

In another preferred embodiment, the mutation(s) is/are point mutation(s), mutation(s) of several amino acids, one or more deletions and/or one or more insertions, and combinations of these mutations in the structural protein, the insertion preferably being the insertion of a cell membrane receptor ligand, a Rep protein or Rep peptide, for example in the form of a Rep domain, an immunosuppressive protein or peptide and/or a

Examples of insertions are, inter alia, integrins, cytokines or receptor-binding domains of cytokines or growth factors such as, for example, GM-CSF, IL-2, IL-12, CD40L, TNF, NGF, PDGF or EGF, single-chain antibodies (scFv) binding to cell surface receptors, for example to single-chain antibodies binding to the surface receptors CD40, CD40L, B7, CD28 or CD34, or epitopes or receptor binding sites which are, for example, in turn recognized by particular antibodies, for example anti-CD40L monoclonal antibodies, or by chemical substances or hormones, for example catecholamines. Further examples are also antibodies against particular epitopes such as, for example, cell recognition particles or parts of xenobiotics such as drugs, which are partly presented on the cell surface of particular cells.

With this indirect targeting it is possible to prepare a universal AAV targeting vector which can be loaded individually, and, depending on the use, with different antibodies, each of which are directed against different specific surface receptors or surface molecules on the target cell, and via which the virus binds to the target cell and is intended to infect the

latter. This makes the individual cloning of different AAV mutants for specific targeting problems unnecessary. It is moreover possible for appropriate vectors or capsid mutants according to the invention
 5 also to be used, by employing a wide variety of antibodies, for determining suitable surface receptors on the target cells which are suitable for virus binding or for uptake thereof into the cells, so that it is possible quickly and efficiently to screen
 10 targeting receptors on the target cells.

It is particularly preferred to insert one or more times - preferably once - the Z domain of protein A, in particular in truncated, deleted form, for example as
 15 Z34C protein (Starovasnik et al. (1997), Proc. Natl. Acad. Sci. USA 16:94, 10080-10085), and, in this case, in some circumstances previously to delete some amino acids at the deletion site in the capsid protein. The Z domain of protein A and successive insertion twice
 20 thereof into the capsid of Sindbis viruses as described by Ohno et al. (1997) Nat. Biotech. 15, 763-767. Protein A binds via five independent domains to the FC part of antibodies. The strongest binding domain is the B or Z domain, of which 33 amino acids are essential
 25 for the binding. This binding structure can be stabilized by two cysteine bridges (Starovasnik et al. supra).

An example of a particularly preferred ligand is the P1
 30 peptide (QAGTFALRGDNPQG) which is a peptide 14 amino acids long from the core sequence of an alpha chain of the laminin family. This sequence is sufficient, for example, to recognize an integrin receptor which mediates, inter alia, the endocytosis of viral
 35 particles, for example of adenovirus. The P1 peptide binds irrespective of its conformation (linear or circular) to the integrin receptor. According to the present invention, the coding DNA sequence of the P1 peptide is inserted into the gene coding for an AAV

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structural protein which is located, for example, on a helper plasmid. Packaging with the mutant helper plasmid results in recombinant AAV with P1 in the capsid (rAAV-P1).

5

Further possible ligands to be inserted at the insertion sites are those which bind merely by their charge, the nature of the characteristic amino acid composition, and/or via their specific glycosylation and/or phosphorylation to cell surface molecules. In this connection, the nature of the characteristic amino acid composition means that these have, for example, predominantly hydrophobic, hydrophilic, sterically bulky, charged amino acid residues or those containing amino, carboxylic acid, SH or OH groups. It is thus possible to make cells susceptible to AAV transfection by a nonspecific mechanism. In this connection, for example, many cell surface molecules are specifically glycosylated and phosphorylated or negatively charged and may thus, for example, represent a target for an AAV mutant with an amino acid ligand with multiple positive charges.

In a further preferred embodiment, the mutation(s) is(are) brought about by insertions at the XhoI cleavage site of the VP1-encoding nucleic acid and in another preferred embodiment at the BsrBI cleavage site of the VP1-encoding nucleic acid. A further preferred embodiment of the structural protein according to the invention is brought about by a deletion between the BsrBI/HindII cleavage sites of the VP1-encoding nucleic acid and one or more insertions, preferably at the deletion site.

In a further preferred embodiment of the present invention, the mutation(s) is(are) brought about by one or more deletions between the XhoI/XhoI cleavage sites of the VP1-encoding nucleic acid, which comprises 62 amino acids (Hermonat, P.L. et al. (1984), J. Virol.,

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51, 329-339). In a further preferred and corresponding embodiment, the deletion(s) is/are located between the BsrBI/HindII cleavage sites of the VP1-encoding nucleic acid, which is located within the deletion described
 5 above and comprises 29 amino acids. This deletion has the advantage that it has no overlap with the rep gene and therefore has essentially no effect on the packaging mechanism.

10 In a further preferred embodiment, one or more insertions are present in the VP3 structural protein (Rutledge, E.A. et al. (1998) supra) before and/or after at least one amino acid in the sequence selected from YKQIS SQSGA, YLTLN NGSQA, YYLSR TNTPS, EEKFF
 15 PQSGV, NPVAT, EQYGS, LQRGN RQAAT, NVDFV VDTNG, because these sites are located on the exposed sites of a loop, in which case the risk of changing the VP3 structure is low.

20 The point mutation(s), the mutation(s) of several amino acids, the deletion(s) or insertion(s) is/are carried out by generally known methods by deletion and insertion in the gene coding for the structural protein. The deletions can be introduced into the
 25 individual structural protein genes for example by PCR-assisted mutagenesis. The insertions can be introduced by generally known methods, for example by hydrolysis by restriction endonucleases of the appropriate structural protein genes and subsequent
 30 ligase reaction.

It is possible relatively easily in an adhesion test (Valsesia-Wittmann, S. et al. (1994) J. Virol. 68, 4609-4619) using suitable cells which express a
 35 selected receptor, for example the laminin alpha receptor, but are difficult to infect with wild-type AAV, for example with wild-type AAV2, to detect the change in the infectivity of the mutated structural proteins according to the invention, for example the

AAV, was more than four orders of magnitude greater than was possible with wild-type AAV.

Another aspect of the present invention is also a structural protein according to the invention in the form of an AAV particle, in particular in the form of an AAV capsid, because particles and capsids are particularly suitable as carriers of selected compounds, for example rAAV transduction vectors.

10 Further aspects of the present invention are a nucleic acid, preferably an RNA or DNA, in particular a double-stranded DNA, coding for a structural protein according to the invention.

15 The present invention also relates to a cell, preferably a mammalian cell, for example a COS cell, Hela cell or 293 cell, comprising a nucleic acid according to the invention. Cells of this type are
20 suitable, for example, for preparing the recombinant AAV particles.

A further aspect of the present invention is therefore also a process for preparing a structural protein according to the invention, in particular for preparing a structural protein according to the invention in the form of an AAV particle, where a suitable cell comprising a nucleic acid coding for the structural protein according to the invention is cultivated and, where appropriate, the expressed structural protein is isolated. For example, the structural protein according to the invention can be isolated on a caesium chloride gradient as described, for example, in Chiorini, J.A. et al. (1995), *supra*.

35 A further aspect of the present invention relates to the use of the fusion protein according to the invention for altering the tropism of AAV, for transforming a cell, in particular a cell whose

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susceptibility to AAV infection was previously low, such as, for example, a haematopoietic cell, for gene therapy in the form of suitable rAAV vectors as already described above in detail, or for genomic targeting.

5

Also included is the use of a fusion protein according to the invention in which the mutation has brought about an increased infectivity for particular cells, for example B16F10 having an integrin receptor, for activity instigations using these cells. Examples are tumour models and tumour cell lines, preferably of murine origin. In this use it is possible to employ for the investigation models which are realistic and comparable for humans and which were not previously accessible in this way, such as certain mouse cell lines. It must be stated in this connection that the susceptibility of mouse cells to infection is generally much worse than that of human cells. Thus, tumours induced in the mouse with B16F10 melanoma cells are not susceptible to AAV2 with unmutated capsid. However, precisely in this case the proteins according to the invention make AAV2 activity studies possible in this and correspondingly other tumour models in mice. An additional facilitating factor is that mouse cells in many tissues and cell types have the specific integrin receptor for the P1 peptide, which is a preferred ligand for the structural proteins mutated according to the invention. It is thus possible with the mutants according to the invention to construct, via the increased infectivity for, for example, B16F10 and other murine tumour cells lines which have, for example, this specific integrin receptor, a test model which is more realistic and more comparable for humans than previously disclosed, because the induced tumours can thus be transduced considerably more efficiently.

35

Another use of the fusion protein according to the invention is in diagnosis. Thus, it is possible according to the invention for example to employ

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antibodies or antibody-binding substances with antibodies as ligands, which recognise and bind particular presented epitopes on cells, for example in a blood sample, so that a signal is initiated.

5 Application examples would be presented parts of xenobiotics such as drugs or cell recognition particles, with which it is possible to determine the origin of tissue cells, for example in tumour diagnosis.

10

Other aspects of the present invention also relate to a medicinal product or a diagnostic aid comprising a fusion protein according to the invention, a nucleic acid according to the invention or a cell according to the invention and, where appropriate, suitable excipients and additives, such as, for example, a physiological saline solution, stabilizers, proteinase inhibitors etc.

15

20 A considerable advantage of the present invention is that through the mutagenesis according to the invention of AAV structural proteins the infectivity can be altered essentially without loss of the packaging efficiency of recombinant AAV vectors into the capsid of the virus, in particular the infectivity for cells of low susceptibility, such as, for example, haematopoietic cells, can be increased several times. The present invention is therefore particularly suitable for an improved in vitro and in vivo transformation of particular cells, for example for somatic gene therapy.

25

30

It will be apparent to those skilled in the art that various modifications and variations can be made to the compositions and processes of this invention. Thus, it is intended that the present invention cover such modifications and variations, provided they come within the scope of the appended claims and their equivalents.

35

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Priority application DE 198 27 457.2, filed June 19, 1998, including the specification, drawings, claims and abstract, is hereby incorporated by reference. All
5 publications cited herein are incorporated in their entireties by reference.

The following examples and figures are intended to explain the invention in detail without restricting it.
10

Fig. 1) shows the detection of P1 on the surface of capsid mutants and the wild type either in a direct ELISA (black bars) or in an indirect ELISA (grey bars).

15 Fig. 2) shows the binding of the capsid mutants or the wild type to various cell types.

Fig. 3) shows the inhibition of the binding of the capsid mutant I-447 to B16F10 cells.
20

Fig. 4) shows the inhibition of the binding of the capsid mutant I-587 to B16F10 cells.

Examples

25 The following mutations were produced by means of PCR-assisted mutagenesis and cutting with the restriction enzymes XhoI, BsrBI and HindIII:

30 1. Mutations in VP1

a) deletion between the XhoI/XhoI cleavage sites of VP-1 (Δ Xho; 62 amino acids, AA) (Hermonat et al. (1984) Journal of Virology 51, 329-339),
35

b) deletion between BsrBI and HindII cleavage sites of VP-1, which is located within the above deletion a) and comprises 29 AAs (Δ BH);

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between different VP proteins so much that packaging does not occur or infection of the target cell is impaired (Table 1).

5 Table 1 Packaging efficiency of the virus mutants prepared

Virus stock	Physical virus titres	Capsid titres (ELISA with A20 MAb)
Wild-type capsid	1.10^{12}	1.10^{11}
VP1 mutants		
Δ Xho	6.10^{12}	5.10^{10}
Δ BH	8.10^{11}	4.10^9
Δ BH+L	1.10^{13}	5.10^{10}
B+L	3.10^{12}	5.10^9
VP3 mutants		
300 (I-447)	1.10^{12}	4.10^{10}
390 (I-534)	1.10^{10}	1.10^7
426 (I-573)	3.10^{10}	1.10^7
440 (I-587)	1.10^{12}	2.10^{10}
565 (I-713)	5.10^{10}	1.10^7

10 The physical virus titres (dot-blot) and capsid titres (A20 capsid ELISA) are shown. The concentrations are stated in particles/ml.

Result:

15 It was possible to show for all 4 VP1 mutants, which are recombinant vectors with LacZ transgene, that mutations do not affect the packaging efficiency, and all mutated viruses can be
20 packaged with good titres similar to those of vectors with unmutated capsid ($\sim 10^{12}$ particles/ml). The AAV vectors with mutations in the VP3 region were also able to be

packaged successfully with LacZ reporter gene (10^{10} - 10^{12} physical particles/ml).

4. Binding of rAAV-P1 to laminin receptor-positive indicator cells

The adhesion tests described above in detail showed that the above mutants infect, in at least one case, the laminin alpha-receptor-positive indicator cells, for example the M07-LP1-R cell line, with an efficiency which is at least 10 times higher than wild-type AAV. It was additionally found in competition assays with soluble P1 peptide that infection with rAAV-P1 is in fact mediated by the inserted ligand.

5. P1 mutation in VP3

The initial starting point was a plasmid pUC-AV2 which was prepared by subcloning of the 4.8 kb BglIII fragment of pAV2 (ATCC 37261, ref. 53) into the BamHI cleavage site of pUC19 (New England BioLabs Inc.). Mutations were carried out at defined sites in the plasmid by means of PCR-assisted mutagenesis known to the skilled person. This entailed a sequence coding for P1, a 14 AA peptide with the AA sequence QAGTFALRGDNPQG, which contains the RGD binding motif of a laminin fragment (Aumailly et al. (1990) FEBS Lett. 262, 82-86), being inserted after nucleotides 3543, 3804, 3921 and 3963. This corresponds to an insertion after amino acids 447, 534, 573 and 587 of the AAV2 capsid protein (named according to the number of amino acids (AA) counted after the AA at the start of the N terminus in VP-1 of AAV2). In the subsequent PCR there is use of in each case 2 mutation-specific primers and, as template, a plasmid, pCap, which contains only the cap gene and is formed by cutting out the 2.2 kb

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The titre resulting from this showed no qualitative difference by comparison with the wild type, although quantitative differences are evident, but they are in turn so small that no domains essential for the packaging can be functionally switched off by the mutations (see Table 2).

It was not possible to read from these results any information about the conformation of the capsid. In a further experiment, A20 monoclonal antibodies (A20MAb) were employed in an ELISA. A20MAb reacts specifically only with completely assembled AAV2 capsid, not with free capsid protein (Wistuba et al., (1997), J. Virol. 71, 1341-1352). The results thereof are also shown in Table 2. Once again, the titre resulting therefrom shows no crucial quantitative or qualitative difference by comparison with the wild type. This shows that the insertions took place on structurally irrelevant loops, and insertion of P1 there had not initiated any change. It was possible to divide the mutations into two groups in relation to their ability of forming DNA-containing particles (Table 2): in one group (mutants I-447 and I-587), the ability to form DNA-containing particles corresponded to the wild-type AAV2. In the second group, this ability was two orders of magnitude less (mutants I-534 and I-573). It was possible to confirm these results by electron microscope analysis.

9. Binding of AAV2 capsid mutants to integrin-expressing cells

The binding of the mutants to the integrin receptor was determined by a cell adhesion assay which was adapted for viral preparations (Aumailly et al., Supra; Valsesia-Wittmann (1994); J. Virol. 68, 4609-4619). 1×10^9 viral particles were coated in 100 μ l of PBS directly onto 96-well microtitre plates and blocked with PBS containing 1% BSA. Controls were coated with a laminin fragment with a concentration of 40 μ g/ml (positive control) or with BSA (10 mg/ml; negative control). 1×10^5 cells per 100 μ l were added to the coated wells. They were incubated at 37° in a humidified incubator for 30 minutes for adhesion. At the end of the adhesion time, the wells were washed twice with PBS in order to remove nonadherent cells. Adherent cells were fixed with 100% ethanol for 10 minutes, stained with crystal violet and quantified by an ELISA reader at 570 nm. This time, B16F10 cells and RN22 cell lines were chosen because they expressed P1-specific integrin on their surface and are resistant to AAV2 infections (Maass, G. et al. (1998) Hum. Gen. Ther., 9, 1049-1059; Aumailly et al., supra). Two of the mutations, I-447 and I-587, bound with similarly great efficiency both to B16F10 and RN22 cells. In distinct contrast to this, there was found to be no binding of the wild-type AAV2 and the mutants I-534 and I-573 to these cells (Fig. 2).

In an inhibition assay, cells were mixed with RGDS or RGES, soluble synthetic peptides in varying concentration, (1-250 μ mol) before they were loaded onto the plate. This experiment was undertaken in order to test the specificity of the binding of the mutants I-447 and I-587 to B16F10 cells. The cell adhesion test was therefore

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carried out in the presence of a peptide (RGDS) which competes for the binding site and which corresponds to the active P1 site, and in the presence of an inactive RGEs peptide. Both mutations I-447 and I-587 were able to bind with 50% efficiency to B16F10 cells with 30 μ mol of the RGDS peptide, whereas the RGEs peptide was inactive even at higher concentrations. At a concentration of 250 μ mol the RGDS peptide completely suppressed virus binding to B16F10 cells (Figs. 3 and 4). Similar results were obtained with RN22 cells.

10. Infection tests with mutants from Example 5

In order to test the tropism of the capsid mutants I-447 and I-587, cell lines Co-115 and B16F10 were infected with the mutated viruses. Co-115 cells were used to test the wild-type receptor tropism of the virions because these cells can be transduced with wild-type AAV2 and do not bind the P1 peptide. The B16F10 cell line was used for the reasons already mentioned in Example 9. Three days after the infection, the cells were investigated by immuno-fluorescence measurement with the aid of an anti-Rep antibody to find whether the viral Rep protein is expressed (Wistuba et al. (1997) J. Virol. 71, 1341-1352; Wistuba et al. (1995) J. Virol. 69, 5311-5319). Cells were cultured on slides to 70% confluence and incubated with various concentrations of viral preparations according to the invention in serum-free medium together with adenovirus 5. The titres of the viral preparations were determined three days later either by in situ detection of Rep protein synthesis in an immuno-fluorescence assay (Rep titre).

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In this case, the immunofluorescence staining with AAV2-infected cells was carried out by a method of Wistuba et al. (Wistuba et al. (1997) J. Virol. 71, 1341-1352; Wistuba et al. (1995) J. Virol. 69, 5311-5319). The slides were washed once with PBS, fixed in methanol (5 min, 4°C) and then treated with acetone (5 min, 4°C). The cells were then incubated with the monoclonal antibodies 76-3, which reacts with AAV2 Rep proteins, at room temperature for one hour. This was followed by washing and incubation with a rhodamine-conjugated anti-mouse secondary antibody at a dilution of 1:50 in PBS with 1% BSA for one hour. The titres were calculated from the last limiting dilution of the viral stock solution which had led to fluorescence-positive cells.

Rep-positive CO115 cells were detectable after infection with wild-type AAV2 and with both mutants I-447 and I-587. The infectivity of I-587 and I-447 for CO115 cells was two to three orders of magnitude less than that of the wild type (Table 3). Transfection of B16F10 cells was just as inefficient with I-447 as with wild-type virus (Table 3). In clear contrast to this, rep-positive B16F10 cells can be detected after infection with I-587, the titre of the I-587 virus being determined at 1×10^6 Rep EFU/ml (Table 3).

In order to investigate whether transfection of B16F10 cells by the mutant I-587 was specifically mediated by the interaction between the P1 sequence on the surface of the mutated capsid and the integrin receptor on the surface of the B16F10 cells, the cells were incubated either with the competing RGDS or with the inactive RGEK peptide at concentrations of 200 μ M before infection with the virus. Addition of RGDS peptide neutralized the infectivity of I-587 for B16F10

cells (Table 3), whereas the control peptide RGES had no effect.

Table 3: Virus titres on the cell surface

Virus stock	Titre on CO115 cells	Titre on B16F10 cells	
		- RGDS	+ RGDS
Wild-type capsid	2.10^9	< 1	nd
Mutants			
I-447	1.10^6	< 1	nd
I-587	1.10^7	1.10^6	< 1
rAAV/LacZ	5.10^7	< 1	nd
rAAV(I-587)/LacZ	6.10^5	5.10^4	< 1

5

The titres on the wild type-susceptible CO115 cells and the wild type-resistant B16F10 cells are shown. The titres are expressed for I-447 and I-587 as for the wild type in Rep EFU/ml and for rAAV/LacZ and rAAV(I-587)/LacZ in LacZ EFU/ml. EFU therein means expression-forming units (Expressing Forming Unit) and nd means "not determined".

10

In a supplementary experiment, a competition test was carried out with heparin, a receptor analogue, in order to rule out the infection of B16F10 cells by the mutant I-587 being additionally mediated by the primary receptor heparan sulphate proteoglycan. With B16F10 cells no change in the infectious titre, that is to say the infectivity of I-587, was detectable after addition of heparin. In contrast to this it was possible to block completely infection of CO155 cells on addition of 50 µg of heparin and above per ml of infection medium. It follows from this that the infection takes place independently of heparan sulphate proteoglycan via P1 ligands and the integrin receptor.

20

25

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11. Infection assay of the mutants from Example 5 with galactosidase

5 In another experiment based on Example 10, rAAV vectors were prepared with a LacZ reporter gene and containing either the wild type (rAAV virion) or I-587 (rAAV(I-587)virion). The viral preparations were called rAAV/LacZ and rAAV(I-587)/LacZ and used to infect B16F10 and
10 CO115 cells (controls).

15 Infected cells were tested for β -galactosidase expression by X-Gal staining three days after the infection. The X-Gal in situ test for cytochemical staining (LacZ titre) was used in this case. According to this, in order to test the expression of β -galactosidase, the cells were washed once in PBS and fixed with 1.5% glutaraldehyde. The cells
20 were subsequently treated with X-Gal (5-bromo-4-chloro-3-indolyl- β -D-galactopyranoside) as already described by Chiorini et al. (1995) Hum. Gen. Ther. 6, 1531-1541. The titres were calculated from the last limiting dilution of the viral stock solution which led to β -galactosidase-
25 producing cells.

Both virions were infectious in the controls on CO115 cells, although the efficiency of rAAV (I-587)/LacZ was 2 orders of magnitude less. With
30 type B16F10 - as expected - no β -galactosidase-positive cells were found after infection with rAAV/LacZ. On the other hand, after infection with rAAV(I-587)/LacZ there were surprisingly found to be a distinctly large number of β -galactosidase-
35 positive cells. The titre of rAAV-(I-587)/LacZ was determined as 5×10^4 LacZ EFU per ml. The infectivity of rAAV vectors for B16F10 cells was improved by more than four orders of magnitude by the mutation according to the invention (Table 3).

12. Z34C protein A mutation in VP3

20 Various mutations in VP3 were carried out in
analogy to Example 5, at the sites mentioned
therein, inserting a sequence coding for the Z34C
domain of protein A (Starovasnik 1997 supra) after
nucleotides 3543, 3804, 3921 and 3963. At the same
25 time, one or more amino acids located at the
insertion site were deleted in each case in order
to avoid problems from too long insertions. The
mutants are prepared as already detailed in
Example 5. Corresponding AAV2 particles were then
30 prepared by the same procedure as in Example 6.

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PATENT CLAIMS

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1. Structural protein of adeno-associated virus (AAV), which comprises at least one mutation, characterized in that the mutated structural protein is capable of particles formation, and the
10 mutation brings about an increase in the infectivity of the virus.

2. Structural protein according to Claim 1, characterized in that the mutation(s) is/are
15 located on the virus surface.

3. Structural protein according to claim 1, characterized in that the mutation(s) is/are located at the N terminus of the structural
20 protein.

4. Structural protein according to claim 1, characterized in that the mutated structural protein brings about a change in the protein-cell
25 membrane receptor interaction.

5. Structural protein according to Claim 4, characterized in that the cell membrane receptor is a glycoprotein of about 150 kD and/or a heparan
30 sulphate proteoglycan.

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6. Structural protein according to claim 1,
characterized in that it is selected from mutated
VP1, mutated VP2 and/or mutated VP3.
- 5 7. Structural protein according to claim 1,
characterized in that it is derived from AAV2,
AAV3, AAV4, AAV5 and/or AAV6.
8. Structural protein according to claim 1,
10 characterized in that the mutation(s) is/are point
mutation(s), mutation(s) of several amino acids,
one or more deletions and/or one or more
insertions, or a combination of this mutation.
- 15 9. Structural protein according to Claim 8,
characterized in that the insertion is a cell
membrane receptor ligand, a Rep protein or Rep
peptide, an immunosuppressive protein or peptide
and/or a protein or peptide having a signal for
20 double-strand synthesis of the foreign gene.
10. Structural protein according to Claim 9,
characterized in that the ligand is selected from
an integrin, a cytokine or a receptor-binding
25 domain of a cytokine, integrin or growth factor, a
single-chain antibody binding to a cell surface
receptor, an antibody against cell surface
structures, an antibody-binding structure or an
epitope, and from ligands which bind via their
30 charge, the nature of the characteristic amino

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acid composition and/or via their specific glycosilation and/or phosphorylation to cell surface molecules.

- 5 11. Structural protein according to claim 8,
characterized in that the mutation(s) is/are
brought about by one or insertions at the XhoI
cleavage site of the VP1-encoding nucleic acid.
- 10 12. Structural protein according to claim 8,
characterized in that the mutation(s) is/are
brought about by one or insertions at the BsrBI
cleavage site of the VP1-encoding nucleic acid.
- 15 13. Structural protein according to claim 8,
characterized in that the mutation(s) is/are
brought about by one or more deletions between the
BsrBi/HindII cleavage sites of the VP1-encoding
nucleic acid and one or more insertions.
- 20
14. Structural protein according to claim 8,
characterized in that one or more insertions in
VP3 is/are located before and/or after at least
one amino acid in the sequence selected from YKQIS
25 SQSGA, YLTLN NGSQA, YYLSR TNTPS, EEKFF PQSGV,
NPVAT, EQYGS, LQRGN RQAAT, NVDFI VDTNG.
15. Structural protein according to Claim 8,
characterized in that the mutation(s) is/are
30 brought about by one or more deletions between

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XhoI/XhoI cleavage sites of the VP1-encoding nucleic acid.

16. Structural protein according to Claim 8,
5 characterized in that the mutation(s) is/are brought about by one or more deletions between BsrBI/HindII cleavage sites of the VP1-encoding nucleic acid.
- 10 17. Structural protein according to claim 1, in the form of an AAV particle, in particular in the form of an AAV capsid.
- 15 18. Nucleic acid coding for a structural protein according to claim 1.
19. Cell comprising a nucleic acid according to Claim 18.
- 20 20. Process for the preparation of a structural protein according to claim 1, characterized in that a cell according to Claim 19 is cultivated and, where appropriate, the expressed structural protein is isolated.
- 25 21. Medicinal product comprising a structural protein according to claim 1.

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22. Medicinal product comprising a nucleic acid
according to Claim 18.
23. Medicinal product comprising a cell according to
5 Claim 19.
24. Diagnostic aid comprising a structural protein
according to claim 1.
- 10 25. Diagnostic aid comprising a nucleic acid according
to Claim 18.
26. Diagnostic aid comprising a cell according to
Claim 19.
- 15 27. Method of using a structural protein according to
claim 1 wherein the method is selected from the
group consisting of altering the tropism of AAV,
transforming a cell, diagnosis, activity
20 investigations, gene therapy, and genomic
targeting.

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534 Rec'd FCT/PTO 18 DEC 2000

PCT/EP99/04288

25 July 2000

MedGene AG

M2689PC BÖ/E1/dor

PATENT CLAIMS

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1. Structural protein of adeno-associated virus (AAV), which comprises at least one mutation, characterized in that the mutated structural protein is capable of particles formation, and the mutation brings about an increase in the infectivity of the virus.

10

2. Structural protein according to Claim 1, characterized in that the mutation(s) is/are located on the virus surface.

15

3. Structural protein according to either of Claims 1 or 2, characterized in that the mutation(s) is/are located at the N terminus of the structural protein.

20

4. Structural protein according to any of Claims 1 to 3, characterized in that the mutated structural protein brings about a change in the protein-cell membrane receptor interaction.

25

5. Structural protein according to Claim 4, characterized in that the cell membrane receptor is a glycoprotein of about 150 kD and/or a heparan sulphate proteoglycan.

30

6. Structural protein according to any of Claims 1 to 5, characterized in that it is selected from mutated VP1, mutated VP2 and/or mutated VP3.

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7. Structural protein according to any of Claims 1 to 6, characterized in that it is derived from AAV2, AAV3, AAV4, AAV5 and/or AAV6.
- 5 8. Structural protein according to any of Claims 1 to 7, characterized in that the mutation(s) is/are point mutation(s), mutation(s) of several amino acids, one or more deletions and/or one or more insertions, or a combination of this mutation.
- 10 9. Structural protein according to Claim 8, characterized in that the insertion is a cell membrane receptor ligand, a Rep protein or Rep peptide, an immunosuppressive protein or peptide and/or a protein or peptide having a signal for double-strand synthesis of the foreign gene.
- 15 10. Structural protein according to Claim 9, characterized in that the ligand is selected from an integrin, a cytokine or a receptor-binding domain of a cytokine, integrin or growth factor, a single-chain antibody binding to a cell surface receptor, an antibody against cell surface structures, an antibody-binding structure or an epitope, and from ligands which bind via their charge, the nature of the characteristic amino acid composition and/or via their specific glycosilation and/or phosphorylation to cell surface molecules.
- 20 11. Structural protein according to any of Claims 8 to 10, characterized in that the mutation(s) is/are brought about by one or insertions at the XhoI cleavage site of the VP1-encoding nucleic acid.
- 25 12. Structural protein according to any of Claims 8 to 10, characterized in that the mutation(s) is/are
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brought about by one or insertions at the BsrBI cleavage site of the VP1-encoding nucleic acid.

13. Structural protein according to any of Claims 8 to 10, characterized in that the mutation(s) is/are brought about by one or more deletions between the BsrBi/HindII cleavage sites of the VP1-encoding nucleic acid and one or more insertions.
14. Structural protein according to any of Claims 8 to 10, characterized in that one or more insertions in VP3 is/are located before and/or after at least one amino acid in the sequence selected from YKQIS SQSGA, YLTLN NGSQA, YYLSR TNTPS, EEKFF PQSGV, NPVAT, EQYGS, LQRGN RQAAT, NVDFT VDTNG.
15. Structural protein according to Claim 8, characterized in that the mutation(s) is/are brought about by one or more deletions between XhoI/XhoI cleavage sites of the VP1-encoding nucleic acid.
16. Structural protein according to Claim 8, characterized in that the mutation(s) is/are brought about by one or more deletions between BsrBI/HindII cleavage sites of the VP1-encoding nucleic acid.
17. Structural protein according to any of Claims 1 to 16 in the form of an AAV particle, in particular in the form of an AAV capsid.
18. Nucleic acid coding for a structural protein according to any of Claims 1 to 16.
19. Cell comprising a nucleic acid according to Claim 18.

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20. Process for the preparation of a structural protein according to any of Claims 1 to 16, characterized in that a cell according to Claim 19 is cultivated and, where appropriate, the expressed structural protein is isolated. .
- 5
21. Medicinal product comprising a structural protein according to any of Claims 1 to 17, a nucleic acid according to Claim 18 and/or a cell according to Claim 19.
- 10
22. Diagnostic aid comprising a structural protein according to any of Claims 1 to 17, a nucleic acid according to Claim 18 and/or a cell according to Claim 19.
- 15
23. Use of a structural protein according to any of Claims 1 to 17 for altering the tropism of AAV, for transforming a cell, for diagnosis, for activity investigations, for gene therapy, and/or for genomic targeting.
- 20

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534 Rec'd PCT/PIO 18 DEC 2000

ABSTRACT

The invention relates to a structural protein of adeno-associated virus (AAV) which comprises at least one mutation which brings about an increase in the infectivity of the virus.

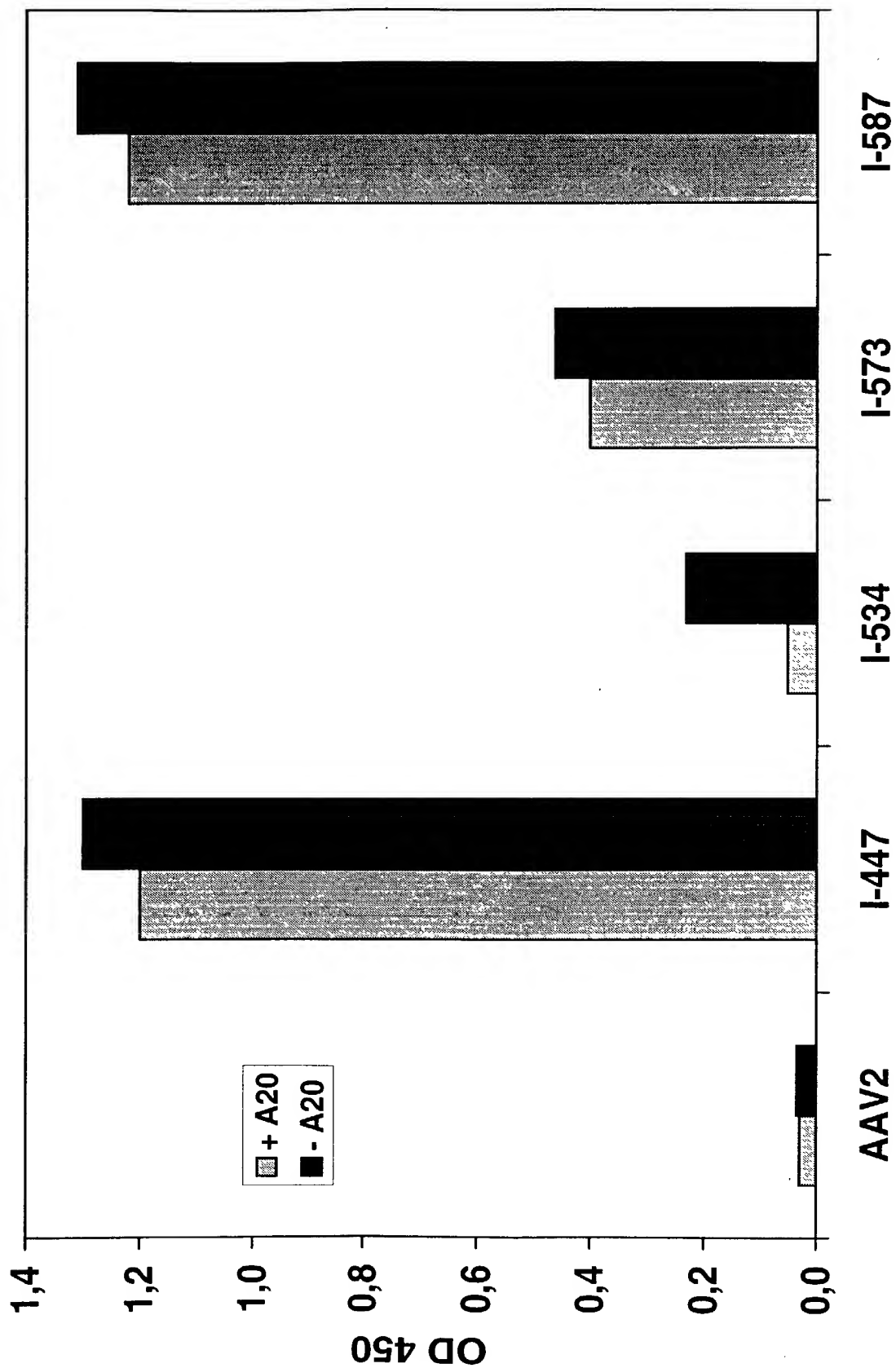


Fig. 1

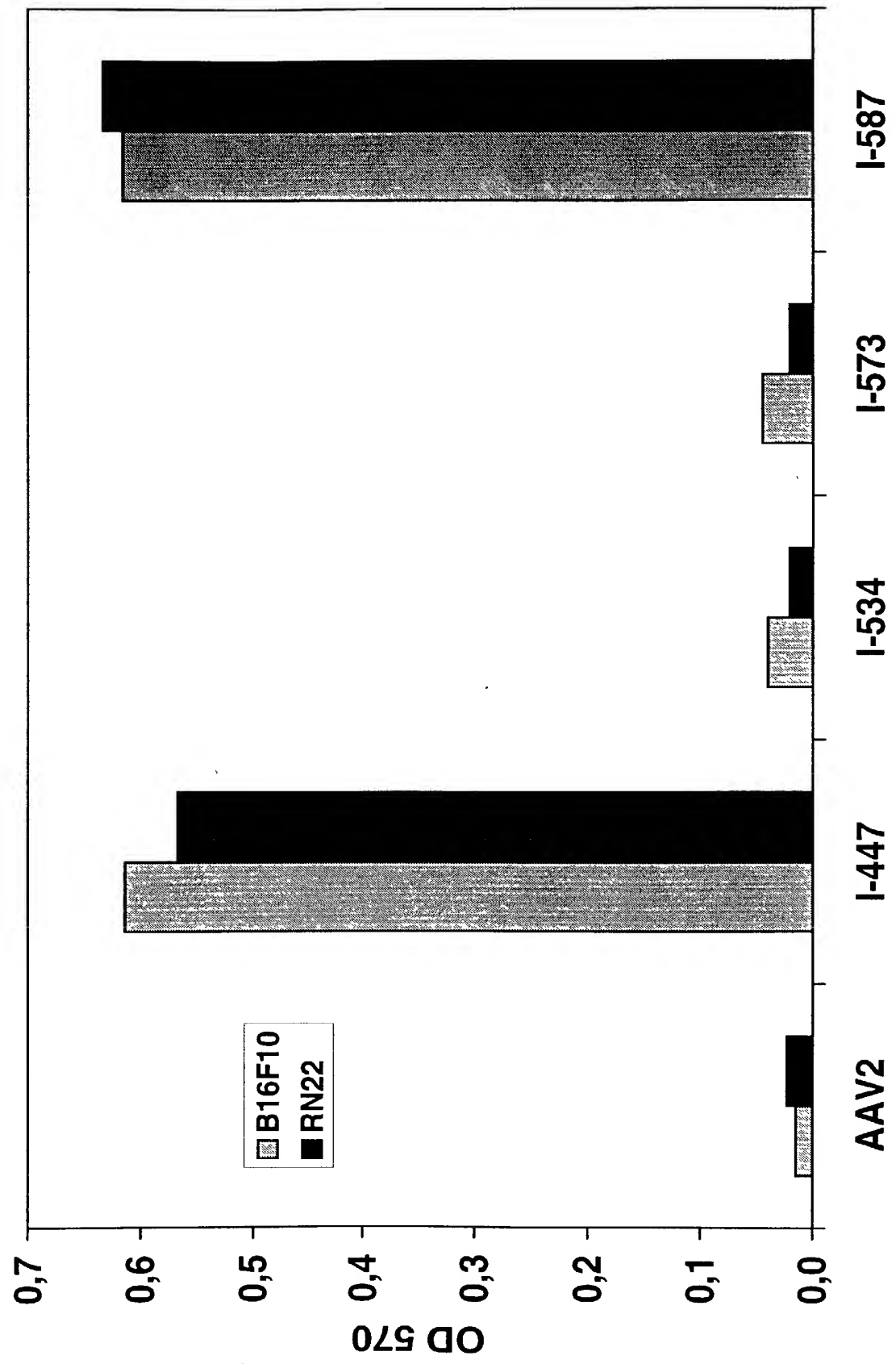


Fig. 2

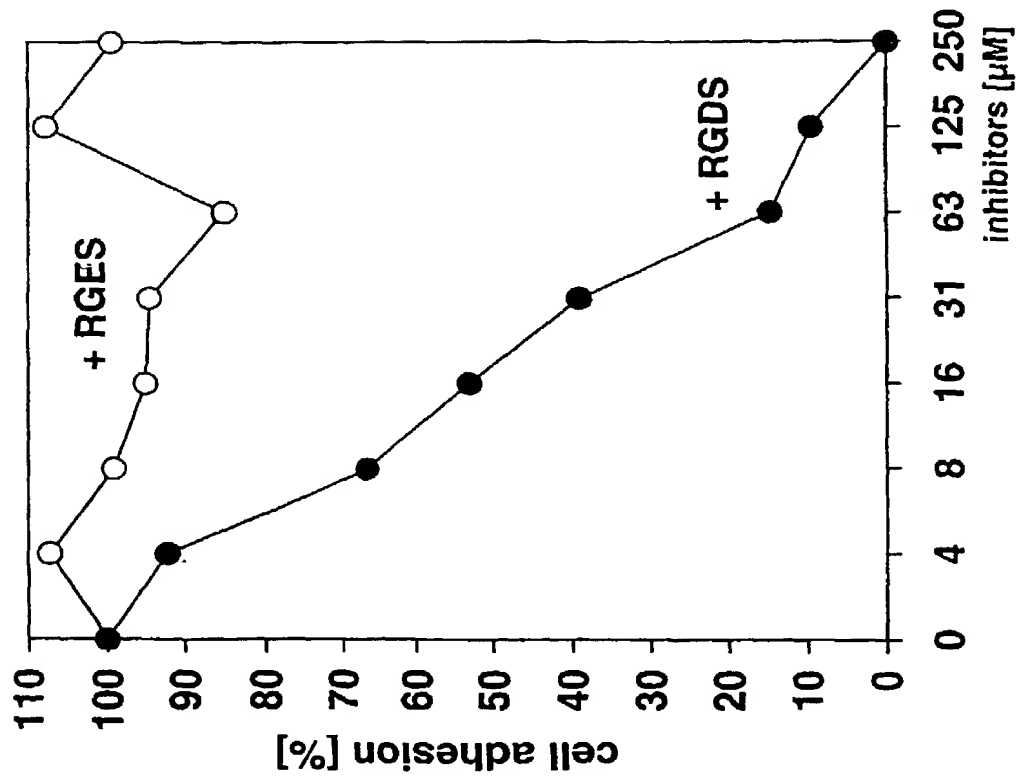


Fig. 3

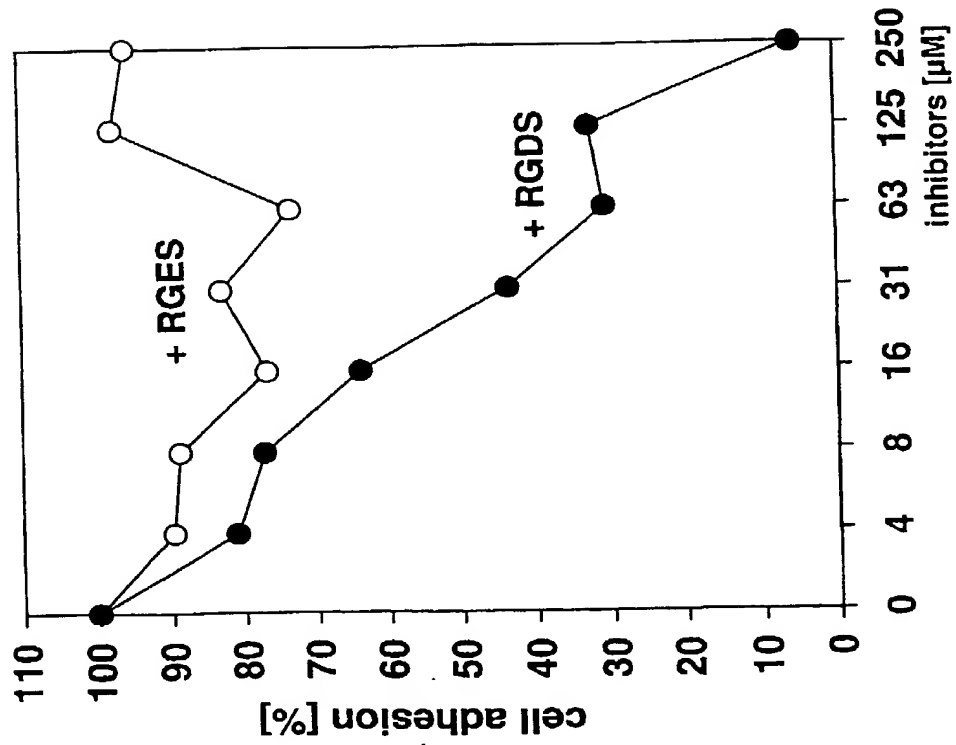


Fig. 4

PATENT
ATTORNEY DOCKET NO: 50125/019001

COMBINED DECLARATION AND POWER OF ATTORNEY

As a below named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled AAV STRUCTURAL PROTEIN, ITS PREPARATION AND USE the specification of which

☐ is attached hereto.

☒ was filed on December 18, 2001 as Application Serial No. 09/720,066
and was amended on _____.

☐ was described and claimed in PCT International Application No. _____
filed on _____ and as amended under PCT Article 19 on _____.

I hereby state that I have reviewed and understand the contents of the above-identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose all information I know to be material to patentability in accordance with Title 37, Code of Federal Regulations, §1.56.

FOREIGN PRIORITY RIGHTS: I hereby claim foreign priority benefits under Title 35, United States Code, §119 of any foreign application(s) for patent or inventor's certificate or of any PCT international application(s) designating at least one country other than the United States of America listed below and have also identified below any foreign application for patent or inventor's certificate or any PCT international application(s) designating at least one country other than the United States of America filed by me on the same subject matter having a filing date before that of the application(s) of which priority is claimed:

Country	Serial Number	Filing Date	Priority Claimed?
Germany	DE 19827457 2	June 19, 1998	Yes
PCT	PCT/EP99/04288	June 21, 1999	Yes

PROVISIONAL PRIORITY RIGHTS: I hereby claim priority benefits under Title 35, United States Code, §119(e) and §120 of any United States provisional patent application(s) listed below filed by an inventor or inventors on the same subject matter as the present application and having a filing date before that of the application(s) of which priority is claimed:

Serial Number	Filing Date	Status

COMBINED DECLARATION AND POWER OF ATTORNEY

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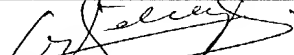
Full Name (First, Middle, Last)	Residence Address (City, State, Country)	Post Office Address (Street, City, State, Country)	Citizenship
<u>Martin Ried</u>	Am Lohwald 36, 86697 Sinning GERMANY DEX	Am Lohwald 36, 86697 Sinning GERMANY	German
Signature: <i>Martin Ried</i>			Date: <i>Sept 09/2001</i>

Full Name (First, Middle, Last)	Residence Address (City, State, Country)	Post Office Address (Street, City, State, Country)	Citizenship
Gilbert Deleage	43, Rue Saint Romain 69008 Lyon FRANCE	43, Rue Saint Romain 69008 Lyon FRANCE	French
Signature:			Date:

Full Name (First, Middle, Last)	Residence Address (City, State, Country)	Post Office Address (Street, City, State, Country)	Citizenship
Anne Girod	Appenzeller Str. 123, 81475 München GERMANY	Appenzeller Str. 123, 81475 München GERMANY	German
Signature:			Date:

[illegible]

Full Name (First, Middle, Last)	Residence Address (City, State, Country)	Post Office Address (Street, City, State, Country)	Citizenship
Martin Ried	Am Lohwald 36, 86697 Sinning GERMANY	Am Lohwald 36, 86697 Sinning GERMANY	German
Signature:			Date:

Full Name (First, Middle, Last)	Residence Address (City, State, Country)	Post Office Address (Street, City, State, Country)	Citizenship
Gilbert Deleage	43, Rue Saint Romain 69008 Lyon FRANCE	43, Rue Saint Romain 69008 Lyon FRANCE	French
Signature: 			Date: 06/09/2001

Full Name (First, Middle, Last)	Residence Address (City, State, Country)	Post Office Address (Street, City, State, Country)	Citizenship
Anne Girod	Appenzeller Str. 123, 81475 München GERMANY	Appenzeller Str. 123, 81475 München GERMANY	German
Signature:			Date:

13.9.2001

COMBINED DECLARATION AND POWER OF ATTORNEY

Full Name (First, Middle, Last)	Residence Address (City, State, Country)	Post Office Address (Street, City, State, Country)	Citizenship
Martin Ried	Am Lohwald 36, 86697 Sinning GERMANY	Am Lohwald 36, 86697 Sinning GERMANY	German
Signature:			Date:

Full Name (First, Middle, Last)	Residence Address (City, State, Country)	Post Office Address (Street, City, State, Country)	Citizenship
Gilbert Deleage	43, Rue Saint Romain 69008 Lyon FRANCE	43, Rue Saint Romain 69008 Lyon FRANCE	French
Signature:			Date:

4-6)

Full Name (First, Middle, Last)	Residence Address (City, State, Country)	Post Office Address (Street, City, State, Country)	Citizenship
Anne Girod	Appenzeller Str. 123, 81475 München GERMANY	Appenzeller Str. 123, 81475 München GERMANY	German French
Signature: <i>[Signature]</i>			Date: 13.9.01

Richard Riemerschmid - Allee 29
81241 München
Germany DEL

PCT/EP99/04288

10.10.1999

MediGene Aktiengesellschaft et al.

M26890PCT BÖ/ps

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